

Four New Alkaloids, Brevianamides O – R, from the Fungus *Aspergillus versicolor*

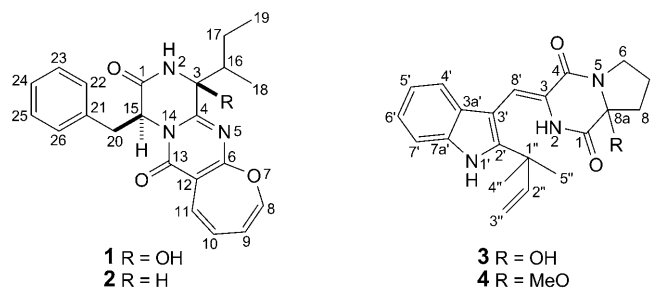
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Four new alkaloids, brevianamides O – R (**1–4**, resp.), were isolated from the AcOEt extract of the solid-state fermented rice culture of the fungus *Aspergillus versicolor*. Their structures were elucidated on the basis of spectroscopic analyses.

Introduction. – *Aspergillus versicolor*, a harmful fungus, is widely found in human and animal foodstuffs. Its secondary metabolites such as aflatoxins and related sterigmatocystins are carcinogenic [1]. Therefore, a variety of secondary metabolites of *A. versicolor* were extensively characterized, including xanthenes such as sterigmatin [2] and sterigmatocystin [3], anthraquinones such as averantin [4], averufin [5], and versicolorone [6], a terpenoid, versiol [7], lactones [8], and chromones such as aspergillitine or aspergiones A–F [9]. Our previous study on the alkaloidal metabolites of the fungus *A. versicolor* led to the identification of five new diketopiperazine alkaloids [10]. The present study disclosed four new diketopiperazine alkaloids, brevianamides O – R (**1–4**, resp.), from the AcOEt extract of the solid-state-fermented rice culture of this fungus. Here, the isolation and structure elucidation of compounds **1–4** are described.



Results and Discussion. – 1. *Structure Elucidation.* Compound **1** was isolated as yellow amorphous powder. The molecular formula ($C_{22}H_{23}N_3O_4$) was determined from the *quasi*-molecular-ion peak at m/z 416.1561 ($[M + Na]^+$) in the HR-ESI-MS. The IR absorption band at $\tilde{\nu}_{\max}$ 3331 cm^{-1} suggested the presence of a OH group. The presence of amides could be deduced from the IR absorption bands at 3167, 1693, and 1658 cm^{-1} ,

and the ^{13}C -NMR signals at $\delta(\text{C})$ 166.1 and 163.2 (*Table 1*). The NMR data and the UV maxima at λ_{max} 209 (4.10), 248 (3.78), and 339 (3.53) nm suggested that compound **1** was an analog of brevianamide L [10]. The comparison of the NMR data of **1** with those of brevianamide L suggested that one O-bearing C-atom resonates at $\delta(\text{C})$ 84.5 in **1** instead of $\delta(\text{C})$ 70.4 in brevianamide L. The OH group must be located at C(3) in view of the key HMBCs of H–C(17) and H–C(18) with $\delta(\text{C})$ 84.5 (C(3)). The HMBC of H–C(16) with C(4), of H–C(10) with C(12), and of H–C(8) and H–C(11) with C(6) suggested the presence of a C=N and a C=C bond between C(4) and N(5), and C(6) and C(12), respectively. The structure of compound **1** was finally elucidated by the HSQC and HMBC experiments.

Table 1. NMR Data of Compounds **1** and **2**. δ in ppm, J in Hz^a)

	1		2	
	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$
C(1)		166.1		167.5
H–N(2)	6.45 (<i>s</i>)		6.50 (<i>s</i>)	
C(3) or H–C(3)		84.5	2.66 (<i>br. s</i>)	58.4
C(4)		156.0		156.1
C(6)		160.6		163.0
H–C(8)	6.06 (<i>d</i> , $J=5.5$)	143.5	6.05 (<i>d</i> , $J=5.6$)	143.2
H–C(9)	5.64 (<i>t</i> , $J=5.5$)	117.2	5.64 (<i>t</i> , $J=5.6$)	117.2
H–C(10)	6.22 (<i>dd</i> , $J=11.2, 5.5$)	128.7	6.18 (<i>dd</i> , $J=11.0, 5.6$)	128.0
H–C(11)	6.80 (<i>d</i> , $J=11.2$)	125.2	6.80 (<i>d</i> , $J=11.0$)	125.5
C(12)		111.3		110.0
C(13)		163.2		160.9
H–C(15)	5.45 (<i>dd</i> , $J=5.6, 3.4$)	57.2	5.47 (<i>t</i> , $J=4.2$)	57.0
H–C(16)	2.36–2.40 (<i>m</i>)	44.2	2.27–2.30 (<i>m</i>)	36.6
CH ₂ (17)	0.83–0.86 (<i>m</i>)	24.6	1.01–1.05 (<i>m</i>)	23.1
Me(18)	0.93 (<i>d</i> , $J=7.2$)	10.7	0.84 (<i>d</i> , $J=7.2$)	15.7
Me(19)	0.79 (<i>t</i> , $J=7.6$)	11.6	0.76 (<i>t</i> , $J=7.6$)	11.9
CH ₂ (20)	3.45 (<i>dd</i> , $J=14.9, 5.6$), 3.55 (<i>dd</i> , $J=14.9, 3.4$)	37.0	3.38–3.41 (<i>m</i>)	37.1
C(21)		134.4		134.3
H–C(22,26)	6.98 (<i>d</i> , $J=7.6$)	130.0	6.94 (<i>d</i> , $J=7.2$)	129.8
H–C(23,25)	7.32 (<i>t</i> , $J=7.6$)	129.3	7.24 (<i>t</i> , $J=7.2$)	128.9
H–C(24)	7.33 (<i>t</i> , $J=7.6$)	128.4	7.30 (<i>t</i> , $J=7.2$)	128.1

^a) Recorded in CDCl₃; ¹H: 600 MHz, ¹³C: 150 MHz; assignments based on HSQC and HMBC.

In the NOESY experiment, the cross-signal between HO–C(3) at $\delta(\text{H})$ 6.84 and H–C(22) or H–C(26) was observed (*Fig. 1*). The hydrolysis of compound **1** in 6N HCl (aq.) for 12 h at 100° afforded L-phenylalanine ($[\alpha]_{\text{D}}^{20} = -34.0$ ($c = 0.1$, H₂O)), which was determined by comparing with an authentic sample. Therefore, the absolute configurations of C(3) and C(15) were determined as (*R*) and (*S*), respectively. The configuration at C(16) could not be determined so far.

Compound **2** was obtained as yellow amorphous powder. The molecular formula (C₂₂H₂₃N₃O₃) was inferred from the *quasi*-molecular-ion peak at m/z 400.1635 ($[M + \text{Na}]^+$) in the HR-ESI-MS. The NMR (*Table 1*) and UV data of compound **2** were

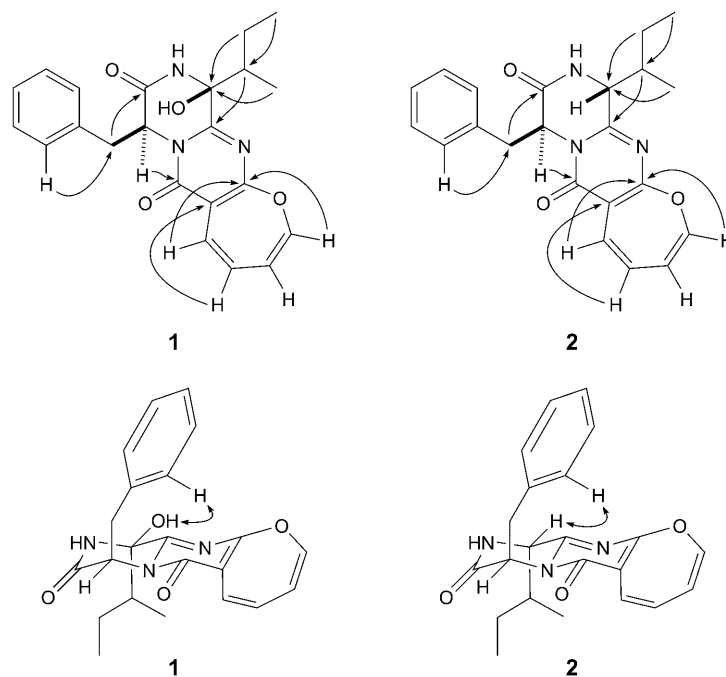


Fig. 1. Key HMBC (H → C) and NOESY (↔) correlations of **1** and **2**

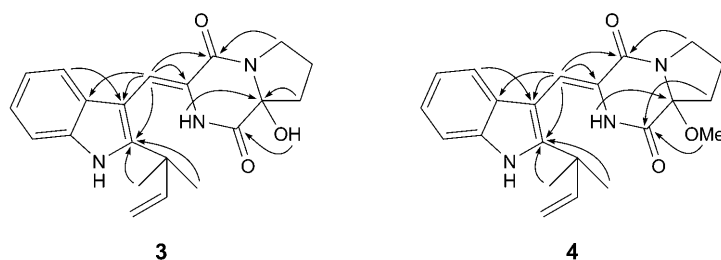
very similar to those of **1**. The molecular formula and IR spectrum suggested that, in compound **2**, there is no HO–C(3) as in compound **1**, which was confirmed by the HMBC of CH₂(17) and Me(18) with $\delta(\text{C})$ 58.4 (C(3)) (Fig. 1). The structure of compound **2** was confirmed by the HSQC and HMBC experiments. The absolute configurations of both C(3) and C(15) were determined as (*S*) on the basis of the NOESY cross-peak between H–C(3) and H–C(22), and of L-phenylalanine obtained from the hydrolysis of compound **2** in the same way as in the case of **1**. Again, the configuration at C(16) could not be determined.

Compound **3** was obtained as a colorless racemic mixture with $[\alpha]_{\text{D}}^{20} = 0.0$ ($c = 0.10$, acetone), and a Cotton effect in the CD spectrum was absent. The quasi-molecular-ion peak at m/z 388.1637 ($[M + \text{Na}]^+$) in the HR-ESI-MS gave the molecular formula C₂₁H₂₃N₃O₃. Signals for 21 H-atoms in ¹H-NMR and 21 signals in ¹³C-NMR spectra were observed (Table 2). The ¹H-NMR signals at $\delta(\text{H})$ 5.09 (*dd*, $J = 11.9, 1.1$, H–C(3'')), 5.12 (*d*, $J = 10.6, 1.1$, H–C(3'')), 6.14 (*dd*, $J = 11.9, 10.6$, H–C(2'')), and the HMBC of $\delta(\text{H})$ 1.56 (*s*, Me(4'')) and 1.57 (*s*, Me(5'')) with $\delta(\text{C})$ 39.2 (C(1'')) and 145.1 (C(2'')) indicated the presence of –Me₂CCH=CH₂ moiety. An *ortho*-substituted benzene ring was evident from the ¹H-NMR signals at $\delta(\text{H})$ 7.43 and 7.37 (*d*, $J = 8.1$, H–C(4'), H–C(7')), and 7.08 and 7.01 (*t*, $J = 8.1$, H–C(6'), H–C(5')). The IR absorptions (1711, 1692 cm^{–1}) and ¹³C-NMR signals ($\delta(\text{C})$ 164.7, 158.8) indicated two amido C=O groups. The above evidence revealed compound **3** as an analog of brevianamide K [10]. The key HMBC of H–N(2) and H–C(7) with $\delta(\text{C})$ 87.2 (C(8a))

Table 2. NMR Data of Compounds **3** and **4**. δ in ppm, J in Hz^{a)}

	3 ^{b)}		4 ^{c)}	
	δ (H)	δ (C)	δ (H)	δ (C)
C(1)		164.7		162.5
H–N(2)	7.99 (s)		7.46 (s)	
C(3)		126.7		125.8
C(4)		158.8		158.7
CH ₂ (6)	3.65–3.69 (m), 3.75–3.77 (m)	44.7	3.72–3.76 (m), 3.89–3.94 (m)	45.3
CH ₂ (7)	2.02–2.04 (m), 2.21–2.27 (m) ^{d)}	19.4	2.00–2.04 (m), 2.13–2.18 (m)	19.2
CH ₂ (8)	2.21–2.27 (m) ^{d)} , 2.21–2.27 (m) ^{d)}	36.0	2.07–2.12 (m), 2.47–2.50 (m)	34.6
C(8a)		87.2		91.7
H–N(1')	10.24 (s)		8.32 (s)	
C(2')		144.2		143.9
C(3')		104.0		103.2
C(3a')		126.7		126.2
H–C(4')	7.43 (d, $J=8.1$)	119.5	7.26 (d, $J=7.2$)	118.8
H–C(5')	7.01 (t, $J=8.1$)	119.7	7.14 (t, $J=7.2$)	121.1
H–C(6')	7.08 (t, $J=8.1$)	121.2	7.19 (t, $J=7.2$)	122.4
H–C(7')	7.37 (d, $J=8.1$)	111.3	7.36 (t, $J=7.2$)	111.3
C(7a')		135.3		134.4
H–C(8')	7.13 (s)	111.1	7.29 (s)	113.0
C(1'')		39.2		39.2
H–C(2'')	6.14 (dd, $J=11.9, 10.6$)	145.1	6.06 (dd, $J=17.3, 10.5$)	144.2
CH ₂ (3'')	5.09 (dd, $J=11.9, 1.1$), 5.12 (dd, $J=10.6, 1.1$)	115.5	5.17 (dd, $J=17.3, 0.8$), 5.21 (dd, $J=10.5, 0.8$)	113.4
Me(4'')	1.56 (s)	27.0	1.52 (s)	27.4
Me(5'')	1.57 (s)	27.2	1.52 (s)	27.3
MeO			3.33 (s)	51.5

^{a)} Assignments based on HSQC and HMBC; ¹H: 600 MHz, ¹³C: 150 MHz. ^{b)} Recorded in CDCl₃. ^{c)} Recorded in (D₆)acetone. ^{d)} Signals overlapped.

Fig. 2. Key HMBC (H → C) correlations of **3** and **4**

indicated the presence of a OH group at C(8a). The structure of compound **3** was finally elucidated by detailed analysis of the HSQC and HMQC spectra (Fig. 2).

Compound **4** was also obtained as a colorless racemic mixture. The molecular formula C₂₂H₂₅N₃O₃ was deduced from the HR-ESI-MS. The IR, UV, and NMR spectra of **4** were similar to those of **3**, but compound **4** contains one more MeO group (δ (H)

3.33 (s, Me), $\delta(\text{C})$ 51.5), which is located at C(8a) in view of the HMBC of $\delta(\text{H})$ 3.33 with $\delta(\text{C})$ 91.7 (C(8a)). The structure of compound **9** was finally determined on the basis of HSQC and HMQC experiments (Fig. 2).

2. *Biological Studies.* Compounds **1–4** were evaluated for their cytotoxic properties against human breast cancer (Bre04), human lung (Lu04), and human neuroma (N04) cell lines. They exhibited no cytotoxicity against these cell lines ($GI_{50} > 10 \mu\text{g/ml}$).

Experimental Part

General. All solvents including petroleum ether (PE; 60–90°) were distilled prior to use. Column chromatography (CC): silica gel (SiO_2 , 200–300 mesh; Qingdao Marine Chemical Factory, Qingdao, P. R. China); Lichroprep C_{18} reversed-phase (RP-18) SiO_2 (40–63 μm , Phenomenex Co.). TLC: SiO_2 GF₂₅₄ (10–40 μm ; Qingdao Marine Chemical Factory, Qingdao, P. R. China). M.p.: X-6 precise melting-point apparatus (Beijing Fukai Science and Technology Development Limited Company). Optical rotations: Perkin-Elmer 341 polarimeter. UV Spectra: Perkin-Elmer Lambda 35 UV/VIS spectrometer; λ_{max} (log ϵ) in nm. IR Spectra: Perkin-Elmer Spectrum One FT-IR spectrometer, KBr pellets; in cm^{-1} . 1D- and 2D-NMR Spectra: Bruker AM-600 spectrometer; δ in ppm rel. to Me_4Si (= 0 ppm), J in Hz. ESI- and HR-ESI-MS: BioTOF-Q mass spectrometer; in m/z (rel. %).

Microorganism and Fermentation. *A. versicolor* (A. 3.3968) was obtained from Institute of Microbiology of the Chinese Academy of Sciences (CAS). It was maintained on potato dextrose agar slant (PDA) at 4° and was stocked in Chengdu Institute of Biology, Chinese Academy of Sciences. The seed culture medium was comprised of dextrose (20 g/l), yeast extract (1 g/l), KH_2PO_4 (3 g/l), $\text{MgSO}_4 \cdot 7 \text{H}_2\text{O}$ (1.5 g/l), and potato extract prepared by extracting 200 g of potato with 1 l of boiling H_2O for 20 min. The pH of the medium was adjusted to 6.0 with 1M NaOH (aq.). The solid culture medium was comprised of rice and 0.3% peptone. The sterilization was carried out at 121° under 15 psi for 30 min.

The fresh mycelium grown on PDA slant at 28° for 3 d was inoculated into 500-ml flasks containing 100 ml of sterilized seed medium. Flasks with inoculated medium were placed in a rotary shaker at 28° and incubated at 180 rpm for 2 d. The seed culture was inoculated into the sterilized solid medium for further fermentation at 28° for 25 d.

Extraction and Isolation. The fermented solid medium (4 kg) was soaked with AcOEt (8 l \times 1, 3 d) at r.t. The solvent was evaporated *in vacuo* to afford a residue (42.0 g). This residue was separated over a SiO_2 column (600 g, \varnothing 90 mm \times 330 mm) eluted with PE/acetone (15:1, 10:1, 5:1, 3:1, 1:1, 0:1; each 3000 ml) to yield five fractions: A (18.6 g), B (5.1 g), C (4.3 g), D (6.0 g), and E (5.0 g). Fr. D was subjected to CC (SiO_2 (110 g, \varnothing 50 mm \times 110 mm); $\text{CHCl}_3/\text{MeOH}$ 30:1, 20:1, 10:1; each 1000 ml) to afford Frs. DA (0.80 g), DB (2.55 g), and DC (1.50 g). Fr. DA was subjected to CC (RP-18 silica gel (22 mm \times 260 mm); MeOH/ H_2O 7:3; 1000 ml) to afford compound **2** (18 mg). Fr. DB was separated by CC (SiO_2 (145 g, \varnothing 44 mm \times 185 mm); PE/AcOEt 2:1, 1:1; each 1000 ml) to give **3** (52 mg), **4** (45 mg), and **1** (12 mg) successively.

Brevianamide O (= (8*S*,11*R*)-8-Benzyl-11-(butan-2-yl)-10,11-dihydro-11-hydroxy-6H-oxepino[2,3-d]pyrazino[1,2-a]pyrimidine-6,9(8H)-dione; **1**): Yellow amorphous powder. $[\alpha]_{\text{D}}^{20} = -125.5$ ($c = 0.10$, acetone). UV (MeOH): 209 (4.10), 248 (3.78), 339 (3.53). IR (KBr): 3331, 3167, 3059, 2963, 2925, 1693, 1658, 1624, 1595, 1427, 1396, 1040, 743. ^1H - and ^{13}C -NMR: Table 1. HR-ESI-MS (pos.): 416.1561 ($[M + \text{Na}]^+$, $\text{C}_{22}\text{H}_{23}\text{N}_3\text{NaO}_4^+$; calc. 416.1581).

Brevianamide P (= (8*S*,11*S*)-8-Benzyl-11-(butan-2-yl)-10,11-dihydro-6H-oxepino[2,3-d]pyrazino[1,2-a]pyrimidine-6,9(8H)-dione; **2**): Yellow amorphous powder. $[\alpha]_{\text{D}}^{20} = -272.3$ ($c = 0.13$, acetone). UV (MeOH): 208 (4.04), 245 (3.68), 336 (3.49). IR (KBr): 3204, 2925, 2854, 1687, 1577, 1535, 1435, 1101, 747. ^1H - and ^{13}C -NMR: Table 1. HR-ESI-MS (pos.): 400.1635 ($[M + \text{Na}]^+$, $\text{C}_{22}\text{H}_{23}\text{N}_3\text{NaO}_3^+$; calc. 400.1632).

Brevianamide Q (= (3*Z*)-8a-Hydroxy-3-[[2-(2-methylbut-3-en-2-yl)-1*H*-indol-3-yl]methylidene]hexahydropyrrolo[1,2-a]pyrazine-1,4-dione; **3**): Colorless crystals. M.p. 184–186°. $[\alpha]_{\text{D}}^{20} = 0.0$ ($c = 0.10$, acetone). UV (MeOH): 204 (4.42), 224 (4.57), 347 (4.20). IR (KBr): 3345, 3268, 2970, 2927, 1711, 1692,

1658, 1576, 1434, 1384, 764. ¹H- and ¹³C-NMR: *Table 2*. HR-ESI-MS (pos.): 388.1637 ($[M + Na]^+$, C₂₁H₂₃N₃NaO₃⁺; calc. 388.1632).

Brevianamide R (= (3*Z*)-8*a*-Methoxy-3-[[2-(2-methylbut-3-en-2-yl)-1*H*-indol-3-yl]methylidene]hexahydropyrrolo[1,2-*a*]pyrazine-1,4-dione; **4**): Colorless crystals. M.p. 206–208°. $[\alpha]_D^{20} = 0.0$ (*c* = 0.10, acetone). UV (MeOH): 203 (4.30), 224 (4.45), 350 (4.04). IR (KBr): 3328, 3166, 3058, 2964, 1693, 1658, 1624, 1600, 1427, 1396, 1040, 743. ¹H- and ¹³C-NMR: *Table 2*. HR-ESI-MS (pos.): 402.1802 ($[M + Na]^+$, C₂₂H₂₅N₃NaO₃⁺; calc. 402.1788).

Cytotoxicity Assay. Cancer cell lines Bre04 (MDA-MB-231), Lu04 (NCI-H460), and N04 (SF-268) were obtained from the *American Type Culture Collection* (ATCC) and cultured according to the supplier's instruction. The cells were seeded into 96-well plates, incubated for 16 h at 37°, and treated with compounds **1–4** at different concentrations for 48 h. Taxol was used as positive control. The cytotoxic activities were examined by means of colorimetric chemosensitivity assay with SRB (sulforhodamine B). The *GI*₅₀ value (the drug concentration required to inhibit the cell growth by 50%) was used as a parameter for cytotoxicity [11][12].

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